

Research article

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A new genus and species, *Brixiola perinetana* gen. et sp. nov., of planthoppers (Hemiptera: Fulgoromorpha: Cixiidae) from Madagascar

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Abstract. A new genus from Madagascar, *Brixiola* Walczak, Gębicki & Taszakowski gen. nov., with its type species *Brixiola perinetana* Walczak, Gębicki & Taszakowski sp. nov. are described and placed in the tribe Brixiini (Hemiptera: Fulgoromorpha: Cixiidae). In its external structure, the new taxon is most similar to species of the genus *Brixia* Stål. Photographs of the adult, illustrations of male genitalia and SEM micrographs of the morphological characters of new species are also provided.

Key words. new genus, new species, planthoppers, Brixiini, taxonomy, Madagascar.

INTRODUCTION

The planthopper family Cixiidae Spinola, 1839 (Hemiptera: Fulgoromorpha) comprises 16 extant tribes (Spinola 1839; Holzinger et al. 2002; Szwedo 2004; Bourgoin 2023). Four of them (i.e., Andini Emeljanov, 2002, Brixidiini Emeljanov, 2002, Brixini Emeljanov, 2002, and Bennini Metcalf, 1938) probably form a genetically close and morphologically similar group (Metcalf 1938; Emeljanov 2002; Ceotto et al. 2008). These tribes, except for the Brixiini, comprise only a few genera and species characterized by forewings held steeply tectiform in repose.

The tribe Brixiini comprises nine genera inhabiting the southern part of the Eastern Hemisphere. The type genus *Brixia* Stål, 1856, which is the largest genus in the Brixiini comprising about 120 species (Stål 1856; Emeljanov 2002), has the broadest range, including the Oriental and Afrotropical regions (Bourgoin 2023) and also the Australasian region, with two known species from Papua New Guinea (Walker 1870). The other genera, with fewer species, show a significant degree of endemism. The richest region in terms of the number of endemics is the Australasian region with the genera *Innobindus* Jacobi, 1928 (seven species), *Solonaima* Kirkaldy, 1906 (15 species), *Undarana* Hoch & Howarth, 1989 (six species), and *Ithma* Fennah, 1969 (three species) (Kirkaldy 1906; Jacobi 1928; Fennah 1969; Hoch & Howarth 1989; Erbe & Hoch 2004; Löcker et al. 2007). A single genus, *Melandeva* Distant, 1906, with two species is known from the Oriental region (Distant 1906; Emeljanov 2007) and there are three genera in the Afrotropical region, namely *Caffrocixius* Fennah, 1967 (six species), *Cibrica* Emeljanov, 2007 (four species), and the monotypic genus *Typhlobrixia* Synave, 1953 (Synave 1952, 1953, 1969; Paulian 1953; Fennah 1967; Van Stalle 1984, 1987; Emeljanov 2007; Soulier-Perkins et al. 2015).

Adults of the tribe Brixiini feed on various plant species, but they are probably not restricted to a specific host plant. Nymphs live underground and feed on the underground parts of plants, like most species of the Cixiidae family; a fact that may explain the tendency to form numerous cavernicolous species (about 30 within the family Cixiidae). These troglobionts feed on the roots of epigeic plants in the subterranean zone of tropical caves, which were formed mainly in sedimentary rocks, and include *Typhlobrixia namorokensis* Synave, 1953 from Madagascar, *Brixia briali* Hoch & Bonfils, 2003 from Réunion, plus six species of the genus *Solonaima* and two species of *Undarana*, which are Australasian endemics (Kirkaldy 1906; Synave 1953; Hoch & Howarth 1989; Hoch et al. 2003; Löcker et al. 2007).

In Madagascar there are 40 Brixiini taxa: 39 representatives of the genus Brixia described so far (Signoret 1860; Jacobi 1917; Lallemand & Synave 1954; Synave 1956, 1965, 1979; Bourgoin 2023) and one endemic troglobiont Typhlobrixia namorokensis (Synave 1952, 1953; Paulian 1953; Soulier-Perkins et al. 2015). Species of the genus Brixia known from Madagascar constitute a large group, but the information about them is fragmentary. The majority of species were reported from the northeastern areas and the eastern coast of the island, also including the areas near the capital city, Antananarivo. Almost all Madagascan species of the genus Brixia are endemics (Signoret 1860; Stål 1866; Jacobi 1917; Lallemand & Synave 1954; Synave 1956, 1965, 1979), except for Brixia krameri Synave, 1979, which also occurs in Seychelles (Holzinger et al. 2008). Brixia species are usually medium-sized, which in Madagascar reach sizes between 5–10 mm in length, and contrastingly coloured. Adult Brixia specimens are usually observed from September to March (Lallemand & Synave 1954; Synave 1956, 1965). Information about host plants of Brixia species found in Madagascar is lacking. The representatives of this genus that occur in nearby Mascarene islands feed on a wide range of plant species belonging to at least 17 genera of 14 families of Magnoliophyta (Williams 1975; Bonfils et al. 1994; Attié et al. 2002, 2005, 2008). Thus, it can be assumed that the trophic preferences of the species of Brixia from Madagascar may be similar to those from Mascarene islands.

Here we described a new cixiid taxa, *Brixiola perinetana* gen. et sp. nov. from Madagascar based on a male collected in Perinet. We owe the description of this new genus and species to Henri Synave (1921–1980) – a Belgian entomologist who described 55 species of the genus *Brixia* Stål, which is closely related to the genus *Brixiola* gen. nov., and over 300 other planthopper species and 25 genera. The label attached by Synave drew our attention and directly helped us to prepare the present study. Synave considered the specimen to belong to the genus *Brixia*.

MATERIALS AND METHODS

Morphological features of the specimen were imaged using a Leica M205C stereo microscope with a Leica LED5000 HDI (high diffuse dome illuminator), a Leica DFC495 digital camera, and the Leica application suite ver. 4.9.0 software. SEM pictures were obtained using Phenom XL field emission scanning electron microscope with a Back-Scatter Detector (BSD). The panoramic image stitcher Image Composite Editor ver. 3.0 and graphic editor Adobe® Photoshop CS6 were used to prepare the figures. The specimen was only cleaned with a brush because other methods, e.g., washing or dehydration, may damage the individual. Next, the specimen was mounted on aluminium stubs (an insect pin was inserted into a piece of the sponge glued to the stub). In the next stage, the specimen was covered with an anti-static spray. The measurements were taken with the Leica application suite ver. 4.9.0 software. The abdomen was boiled three times (about 10 minutes) in a 10% solution of potassium hydroxide (KOH) according to Knight's method (Knight 1965). The genitalia were detached from the abdomen after boiling. The specific parts of the genital structures were separated from the abdomen using thin forceps and a needle blade. Next, these structures were placed in glycerine. The genitalia were studied and drawn with the Nikon Ni-U light microscope with a camera lucida.

The morphological terminology of the head, thorax and legs follows Holzinger et al. (2003) and Ceotto & Bourgoin (2008), for male genitalia it follows Bourgoin (1987), and the terminology of venation of fore- and hindwings follows Anufriev & Emeljanov (1988) and Bourgoin et al. (2015). The terminology and classification of sensilla follow Bourgoin & Deiss (1994), Holzinger et al. (2002), Romani et al. (2009), and Wang et al. (2018).

In the description of the type labels, the contents of each label are enclosed within double quotation marks (""), the individual lines of data are separated by a double forward slash (//), and the author's comments are between square brackets ([]).

RESULTS

Brixiola Walczak, Gębicki & Taszakowski gen. nov. um:lsid:zoobank.org:act:906B4366-62F2-4859-9DE2-CAD2191D073C (Figs 1–4)

Type species: *Brixiola perinetana* Walczak, Gębicki & Taszakowski gen. et sp. nov.

Diagnosis

Apical part of the head elongated and very narrow, its width slightly smaller than width of compound eye. Frons widest much above the median ocellus and considerably tapering towards the frontoclypeal suture. Lateral carinae in the apical part of the head located very close to each other, but not fused. Narrower (apical) part of the frons concave, delimited by two transverse carinae in the upper apical area of the head. Antenna base located above posterior margin of compound eyes. Semicircular scapus four times shorter than cylindrical pedicellus. Median ocellus located at the distance slightly greater than its diameter from the arcuated frontoclypeal suture. Lateral margins of frons and clypeus clearly intended in the area of frontoclypeal suture (these margins do not form one uniform arcuate line). Outer edge of metatibia with a row of small spines. Apex of metatibia with a rim of spines including two long lateral and two short middle spines. Apex of metatarsomere I with one long and three short spines; apex of metatarsomere II with one long and two short spines. The basal concavity of costal margin of tegmen absent. In forewings, ScP considerably delimited the posterior margin of lanceolate pterostigma. Length of common stem of ScP+R+M almost equal to the length of basal cell. Vein R separating in a total of seven branches: RA branches into RA, and RA, and RP branches into $RP_1 - RP_5$ There are five MP branches, initially fused in a common stem MP, which forks into MP₁₊₂ MP₃ and MP_{4+5} branches. CuA₁ arched opposite to MP_{4+5} In the hindwings r-m connected with M at the point where MP separating from M. The length of the common vein Cu almost equal to the length of basal cell (bc). The section of the inner edge located between the apex of clavus and the end of oblique vein icu strongly shortened. Slender periandrium with broad basal crest-like appendage and four long apical spines. Flagellum in the ventral view with a short apical spine and a row of small spines and in the lateral view with three broad short teeth and long basal spine. Apex of parameres pallete-shaped extended. Apical lobes of anal tube narrow and curved downwards.

Description

Head. Head including eves narrower than pronotum. Vertex trapezoidal, approximately two times wider than long, deeply indented in the middle, lateral carinae merged and connected by short arcuate carina. At apex of vertex short groove. Width of basal part of vertex slightly smaller than width of compound eye (in dorsal view) (Fig. 1B). Frons strongly expanded, its lower part distinctly wider than compound eye, oval-shaped. Lateral carinae weakly concave. Apical part of head marginated with two short transverse and separated carinae. Transverse subapical carina interrupted in the middle and strongly arched. Apical carina (separating frons from vertex) oblique. These carinae delimited a small area of irregular shape located in the apical part of the head between vertex and frons. Frontoclypeal suture weakly visible, concave, arched, its convexity directed towards median ocellus. Postclypeus slightly shorter than frons. Anteclypeus at least three times shorter than postclypeus. Both with distinct median carina, reaching about half of the length of postclypeus. Lateral margins of frons and clypeus (in lateral view) considerably intended in the area of frontoclypeal suture (they do not form one uniform arcuate line). Rostrum long, its tip exceeds the line between coxae III (hind coxae) (Figs 1A, 2A). Median

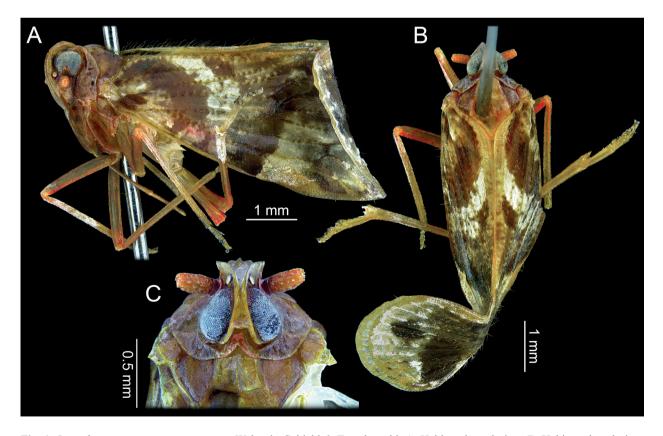


Fig. 1. *Brixiola perinetana* gen. et sp. nov., Walczak, Gębicki & Taszakowski. A. Habitus, lateral view. B. Habitus, dorsal view. C. Vertex and pronotum, dorso-anterior view.

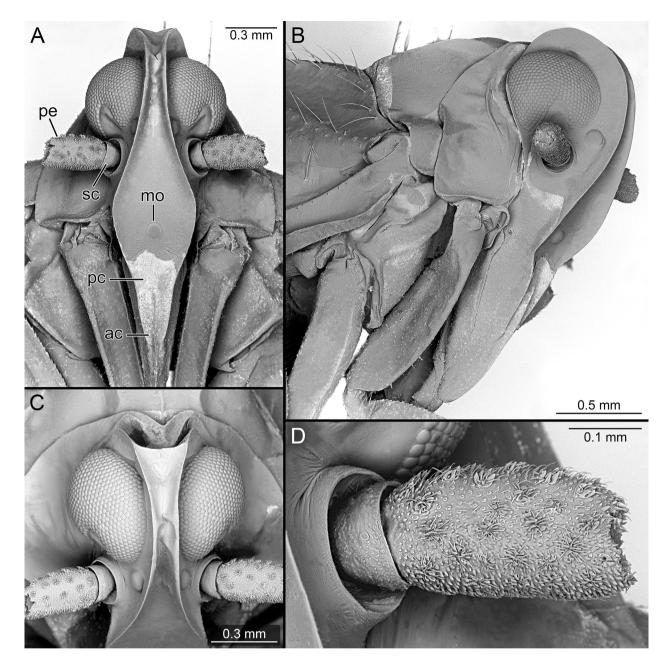


Fig. 2. *Brixiola perinetana* gen. et sp. nov., Walczak, Gębicki & Taszakowski, SEM detail of morphology. A. Head, frontal view. **B**. Head, lateral view. **C**. Vertex and pronotum, anterodorsal view. **D**. Antenna, lateral view. Abbreviations: ac = anteclypeus; mo = median ocelli; pc = postclypeus; pe = pedicellus; sc = scapus.

ocellus distinct, located at the distance slightly greater than its diameter from the arcuated frontoclypeal suture. Lateral carina regularly rounded (Fig. 2A). Above ocelli, near margin of the compound eye, two sockets of sensory organs (Fig. 3C). On gena above the arcuate genal groove, near the hind margin of head, sensory organ with complex structure (Fig. 3B). Cylindrical pedicellus four times longer than semicircular scapus, about twice longer than wide. Antennae embedded in deep cavities highly fringed with cuticular collar (Fig. 2D). Surface of pedicellus (beside narrow basal part) covered with sensory plate organs (Figs 2D, 3A). Centrally located, slightly convex sensory plate covered on the inner side with single ones, and on outside most often by long, curved and tapered processes clustered in pairs. The ventral side of pedicellus covered densely with cuticular processes (also occurring within plate organs). Single sensilla trichoidea (short and straight) are scattered between them. Dorsal side of pedicellus with cuticular microtubercles (much larger than on ventral side) almost exclusively within sensory plate organs. Between plate organs, numerous sensilla trichoidea (long and curved) (Fig. 2D).

Table 1. Comparison	of the Brixiola gen.	nov. and the genus Brixia.
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Feature	Brixiola gen. nov.	Brixia
apical part of the frons (Figs 2A-C)	elongate and concave; median suture absent	flat; short median suture present
basal concavity of forewing costal margin (Fig. 3F)	absent	present
RP veins on forewing (Fig. 3F)	five RP veins, RP_3 and RP_4 connected in the basal part	three to five RP veins, the last two connected in the basal part
apex of metatibia	with seven spines, including two lateral, longest and two median, shortest spines	with six spines, the outer longest, and two median, the shortest spines
periandrium (Figs 4A–B)	with a large ventral crest without spines	without the ventral crest or with ventral crest with spines
flagellum (Figs 4A–B)	with an elongated spine, row of small spines, three short lateral teeth and one apical teeth	only with long spines
apical lobes of anal segment (Figs 4E–F)	narrow and curved downwards	broad and not curved downwards

Thorax. Pronotum very short, with broad paranotal lobes and posterior margin intended in right angles (Figs 1B–C, 2C). Mesonotum with three longitudinal carinae including median carina reaching the base of scutellum (Fig. 1C).

Legs. Fore legs: procoxa elongated, about three times longer than wide, slightly concave on ventral site, with long, arcuate groove. Profemur 1.6 times longer than procoxa, slightly concave on the lateral side, the inner side with median ridge armoured with small denticles. Lower margin with very small spines. Protibia about 1.4 times longer than profemur (Fig. 1A). Middle legs: mesocoxa elongated and widest in basal part with a straight groove on the lateral side. Mesofemur short about 1.4 times shorter than coxa. Mesotibia rectangular in cross-section, about three times longer than mesofemur with margins densely covered with small spines. Second tarsomere shortest. Hind legs: metacoxa robust with spines. Outer margin of metatibia with a row of small spines. Apex of metatibia with a rim of seven spines including two long lateral spines (the inner spine is slightly shorter than outer one), three short median spines connected to each other and two medium-sized spines on inner surface of apex of tibia. The apex of metatarsomere I with one long and three short spines; apex of metatarsomere II with one long and two short spines. All tarsomeres equal in length (Fig. 1A).

Wings. Forewing strongly expanded in the anterior part. Basal and middle band reaching the costal margin of clavus where they are widest. The basal concavity of costal margin of tegmen absent. The common stem of ScP+R+M almost as long as basal cell. RA forks into RA₁ and RA₂. Long stem RP forks behind pterostigma into single RP₁, RP₂, bifurcated RP₃₊₄ and RP₅. MP forks into MP₁ and MP₂, although short stem MP₃₊₄₊₅ forks into single MP₃ and forked MP₄ and MP₅ (according to the

venation pattern: MP₁₊₂ MP₃ and MP₄₊₅). CuA distinctly bent doubly arched. The length of oblique vein icu not greater than that of transverse veins. Forking of CuA, and icu located slightly above the apex of clavus. The section of inner edge between apex of clavus and end of oblique vein icu strongly shortened. Most apical cells are the same size and shape and slightly longer than corresponding subapical cells (Figs 3D-E). Hind wing. Costal margin of hind wings slightly arched. Nodus elongated and narrow, located approximately in the middle of the wing length. Forkss of RP, and RP, reaching the anterior margin of nodus (before the apex of nodus). Relatively long vein r-m connected with M at the point where MP separating from M. Common stem of MP and CuA, elongated and slightly shorter than r-m. The length of the common vein Cu (at the base of the wing) almost equal to the length of basal cell (bc) (Figs 3F-G).

Male genital structure. Aedeagus relatively long and narrow with roundish ventral crest (Fig. 4A). The dorsal edge of periandrium almost straight (without crest), basal lobe directly connected with the anal segment (Fig. 4F). In frontal part of periandrium, a pair of elongated spines with combined bases. First one long, reaching the base of periandrium, second one bent backward in the middle of its length. On both sides of the apical part of the periandrium, one single straight spine (Figs 4A–B). Flagellum in the ventral view with a short apical spine and a row of small spines and in the lateral view with three broad short teeth and long basal spine. Parameres with long stems and obliquely placed apical lobes (Fig. 4C). Anal segment with large anal style. Apical lobes of anal tube narrow and curved downwards (Figs 4D–E).

Remarks. In its external structure, including the male genitalia, *Brixiola* gen. nov. is similar to many representatives of the tribe Brixiini and even Brixidiini. *Brixiola* gen. nov. is distinguished by elongated, laterally flattened

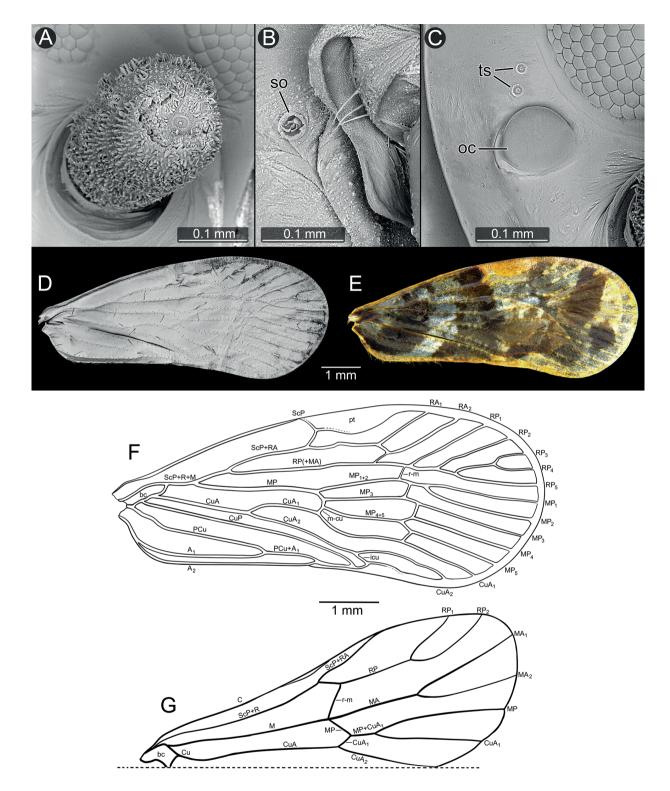


Fig. 3. *Brixiola perinetana* gen. et sp. nov., Walczak, Gębicki & Taszakowski, SEM detail of morphology. **A**. The apical part of the pedicel, top view (flagellum on both antennae broken off). **B**. Sensory organ with a complex structure on gene. **C**. Two sockets of sensory organs above ocelli. **D**. Forewing, **E**. Forewing, photo in colour. **F**. Forewing, drawing. **G**. Hindwing, drawing (visible part). Abbreviations: $A = Anal vein (A_1-A_2)$; bc = basal cell; C = Costa vein; Cu = Cubitus vein; $CuA = Cubitus anterior vein (CuA_1-CuA_2)$; CuP = Cubitus posterior vein; icu, m-cu and r-m = transverse venation; M = Media vein; MA = Media anterior vein; $MP = Media posterior vein (MP_1-MP_5)$; oc = ocelli; PCu = Postcubitus vein; pt = pterostigma; Sc = Subcosta vein; ScP = Subcosta posterior vein; $R = Radius vein (R_1-R_2)$; RA = Radius anterior vein (RA_1-RA_2); RP = Radius posterior vein (RP_1-RP_5); so = sensory organ with complex structure; ts = two sockets of sensory organs.

the genus Brixia (Williams 1975).

vertex and a narrow apical part of frons framed with high

lateral carinae, which differs it from Australian endem-

ics that possess a much shorter head. The width of the

vertex (at its base) clearly exceeds its height in the ge-

nus Innobindus, while in Solonaima the vertex is at least

three times and in Undarana even six times wider than

its height (Kirkaldy 1906; Jacobi 1928; Hoch & Howarth

by wide and flattened frons lacking high lateral carinae,

whereas Typhlobrixia is characterised by the reduction of

compound eves and hypogeic habitat (Synave 1953: Fen-

nah 1967; Van Stalle 1987). The oriental genus Melan-

deva is characterised by different morphological fea-

tures, including a mesonotum with five carinae (there are

three carinae in Brixiola gen. nov.) and forewings with

increased number (7) of MP branches (the first forking

of MP is its division into $MP_{1+2+3+4}$ and MP_{5+6+7} branch-

es) (Distant 1906; Emeljanov 2007). Representatives of

the genera Caffrocixius and Solonaima possess a simi-

larly characteristic structure of the mesonotum (Kirkaldy

1906; Fennah 1967; Van Stalle 1987). The genus *Cibrica* is unique by its forewing venation pattern, with MP fork-

ing into MP_{1+2+3} and MP_{4+5} branches (MP_{1+2} and MP_{3} and MP_{4+5} in *Brixiola* gen. nov.). The head morphology in the

genus *Brixiola* gen. nov. is similar to the genera *Cibrica* and *Brixidia* Haglund, 1899 (Haglund 1899; Holzinger

et al. 2002; Emeljanov 2007). These three genera possess a narrow and laterally flattened head with deeply con-

cave vertex surrounded by raised ridges and frons framed

with foliate lateral carinae not merging in the middle of

the apical part of frons. The genus Brixidia is identified

by apical cells of tegmina considerably longer than sub-

apical cells and CuA and icu (long and oblique) forking

clearly above the apex of the clavus. Species of the genus

Brixia may be identified by the presence of lateral cari-

nae fused in the apical part of frons and forming of the

short median groove between them. The apical part of the

head formed this way has a different length in species of

Small basal concavity of the basal part of the costal

margin of tegmina has been indicated as a characteristic

feature of the genus Brixia (Ceotto & Bourgoin 2008),

but it is also present in other representatives of Cixiini, i.e., *Borbonomyndus* Attié, Bourgoin & Bonfils, 2002

assigned to the tribe Oecleini Muir, 1922 and Eucarpia

Walker, 1857 assigned to the tribe Eucarpiini Emeljanov,

2002 (Walker 1857; Muir 1922; Attié et al. 2002; Emel-

janov 2002; Holzinger et al. 2002; Tsaur & Hsu 2003;

Löcker et al. 2010). Therefore, this feature cannot be

considered as characteristic of the genus Brixia. Basal

M at the point where MP separating from M is present

African species of the genus *Caffrocixius* are clearly distinguished from other genera of the tribe Brixiini,

1989).

tallini), *Andes* Stål (Andini) and *Brixidia* (Brixidiini) of subfamily Cixiinae and *Bothriocera* Burmeister of subfamily Bothriocerinae (Emeljanov 2002). In many representatives of these genera there is a vestigial section of M at the point where r-m is connected to M vein (e.g., in *Pintalia* and *Brixidia*).

The characteristic feature of *Brixiola* gen. nov. is the presence of arched RP_1 and RP_2 reaching the costal margin. Both RP_1 and RP_2 reach apical or distal margin of the hindwing in most representatives of the Cixiinae. In *Brixidia* the venation pattern is similar to that in *Brixiola* gen. nov. with characteristic pattern of radial veins (very short RP_1 is connected with anterior wing margin, while straight RP_2 is connected with apical wing margin). The poor condition of the holotype makes it impossible to describe all features of the hindwings, especially their apical and jugal parts.

The genus *Brixiola* gen. nov. is the most morphologically similar to the genus *Brixia* (including Madagascan species). The features that distinguish these genera are summarized in Table 1.

Brixiola perinetana Walczak, Gębicki & Taszakowski gen. et sp. nov.

urn:lsid:zoobank.org:act:3FB6D23B-58D5-46C9-89EB-1C0C4340F05C (Figs 1–4)

Diagnosis

See generic diagnosis.

Description. Medium-sized species. Measurements of morphological structures are presented in table 1. Vertex trapezoidal. Frontal and lateral carinae do not merge at the top of the head. Frons without spots. Head uniformly dark-yellow. Labrum uniformly light-yellow. Pedicellus dark-yellow. Pronotum uniformly dark-brown, mesonotum with dark sides and the light central disc between lateral carinae, margins darker. Legs pale with dark-yellow, in some places, pink ridges (Figs 1A-B). Wing background hyaline with three brown bands and apical spots. In the forewing, costal margin, pterostigma and inner margin of clavus yellow. Ends of longitudinal veins yellow. The colour pattern of the wing irregularly spotted (Figs 1A, 3E). Between MP_{1+2} and MP_{3+4+5} round, brown spot. The vicinity of basal cell dark-brown. Longitudinal veins with vellowish setae embedded on pale tubercles. On light forewing background, three brown bands and one apical round spot. Hindwing milky-white with dark-brown longitudinal veins (Figs 1A-B, 3E). Sternites paler at the base, with a light-orange middle part and red margins. Ventral crest of periandrium round. Periandrium with four spines, flagellum with one long spine (Figs 4A-B).

Measurements. BODY: total length (with forewings): 8.41. HEAD: Frontal part length (including vertex + frons + clypeus): 2.32, frons length in midline: 1.13, frons total

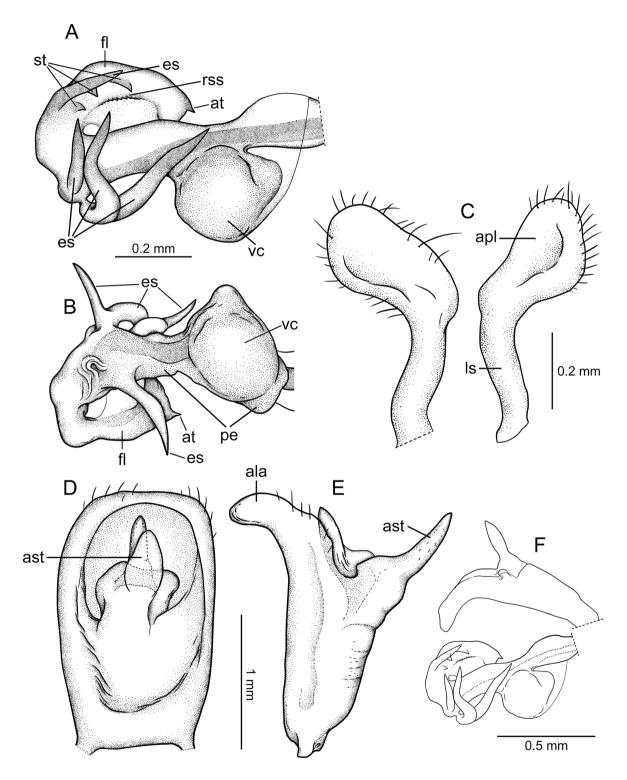


Fig. 4. Brixiola perinetana gen. et sp. nov., Walczak, Gębicki & Taszakowski, male genitalia. A. Aedeagus, side view. B. Aedeagus, ventral view. C. Styles, frontal view. D. Anal segment, dorsal view. E. Anal segment, side view. F. Laying of the anal segment with the aedeagus before preparation. Abbreviations: ala = apical lobe of anal segment; apl = apical lobes of paramere; at = apical teeth; es = elongated spines; fl = flagellum; ls = long stem; ast = anal style; pe = periandrium; rss = row of small spines; vc =, ventral crest of periandrium.

length: 1.41, frons width: 0.50, interocular distance 0.85, vertex in midline: 0.32, vertex total length: 0.39, vertex width: 0.27, eye maximum width: 0.58, clypeus maximum length in midline: 1.07, clypeus maximum width: 0.36, pedicel length: 0.37, pedicel width: 0.21. PRONO-TUM: length in midline: 0.09, total length: 1.47, width: 1.76. MESONOTUM: length in midline: 1.29, width (without tegulae): 1.36. HIND LEG: femur: 1.37, tibia: 3.38, tarsus: 1.59, tarsomere: I: 1.01, tarsomere II: 0.61, tarsomere III: 0.44. FOREWING: length: 7.35, width: 3.05.

Distribution. Madagascar; species known from one locality in the eastern part of the island.

Material examined. *Holotype*, 1♂: "Holotype // Brixiola perinetana // gen. et sp. nov. // Walczak, Gębicki & Taszakowski / det. 2022" [red label] [bold font]; "MADAGASCAR Tam. // Perinet // 28.IX.58 F. Keiser" [bright pinkish label]; "Licht- // fang" [bluish-greenish label]; "♂" [dirty white label]; "H. Synave det. 1964 [64 is hand-written] // Brixia nov. sp." [handwritten] [dirty white label]. Examined material is deposited in the collection of the Naturhistorisches Museum, Basel, Switzerland (NHMB).

Ecology. Unknown.

Etymology. The name refers to Perinet, the place where the specimens were collected. Perinet is situated about 60 km from Antananarivo (Eastern Madagascar, near Moramanga in the vicinity of the Andasibe-Mantadia National Park).

Remarks. Brixiola perinetana gen. et sp. nov. is similar in body colouration to some representatives of the genus Brixia, especially to Brixia trifasciata Synave, 1956. Brixiola perinetana gen. et sp. nov. differs from B. trifasciata in having clearly uniform pale face, dark brown pronotum and a different arrangement of three fuscous stripes on the forewings (Synave 1956). Brixiola perinetana gen. et sp. nov. possesses two thicker anterior stripes, and one of them adheres directly to the pterostigma. The third stripe is only partially developed (reaching only MP) and is located on the apical area of the forewing. Brixiola perinetana gen. et sp. nov. differs from the similar species Brixia pullomaculata Lallemand & Synave, 1954 in having a considerably narrower and straight third stripe not reaching the apex of the forewing and the presence of a fuscous apical spot (Lallemand & Synave 1954).

Key to the African genera of the tribe Brixinii Stål, 1856

The present identification key to the African genera of the tribe Brixinii is based on external morphological characters.

1. Compound eyes reduced, frontal ocellus absent, lateral ocelli partly reduced (diminished). Tegminae with ScP and R forming a common basal stem (ScP+R); MP arises directly from basal cell

- *Typhlobrixia* Synave, 1953 Compound eyes well developed (spherical). Three ocelli present. Tegminae with ScP, R and MP arising
- from basal cell as a common stem (ScP+R+MP) ... 2
 Head broad, posterior part the same width as compound eye (in dorsal view), the length of its dorsal part about equal to the width (at the base of the head). Lateral carinae of vertex and frons not foliate, frons flat and broad, elliptical; long median groove reaching median ocellus. Mesonotum with five carinae (pair of submedian carinae significantly shorter than lateral pair)

- **3.** Lateral carinae in the apex of head completely fused, forming narrow interocular part. Median groove of frons short, not reaching median ocellus *Brixia* Stål, 1856
- 4. Vein MP forking into MP₁₊₂ and MP₃₊₄₊₅. Hind tibiae with short spines...... *Brixiola* gen. nov.
- Vein MP forking into MP₁₊₂₊₃ and MP₄₊₅. Hind tibiae without short spines........... *Cibrica* Emeljanov, 2007

DISCUSSION

Comparative studies on the morphology of species of the tribe Brixiini, especially the species of the genus Brixia, have not been conducted so far. The large genus Brixia exhibits a remarkable morphological diversity. The descriptions of species of the genus Brixia are often not precise, especially those in the old publications. The designation of groups within Brixia is based almost entirely on colouration, particularly wing colouration, which makes it difficult, and in many cases impossible, to confirm or correct the descriptions of many species. The genus *Brixia* needs a thorough revision, and we would not be surprised that after a comprehensive study of the Madagascan taxa, some species currently in Brixia are moved to Brixiola gen. nov. Considering the literature data, it seems possible that the Madagascan species Brixia pullomaculata Synave, 1956 belongs to Brixiola gen. nov., evidenced by the morphology of the frontal part of the head (Synave 1956).

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REFERENCES

- Attié M, Bourgoin T, Bonfils J (2002) The Cixiidae (Hemiptera: Fulgoromorpha) of the Mascarenes islands and Madagascar. Endemism and description of new taxa from Réunion with notes on their host plants. European Journal of Entomology 99: 543–555. https://doi.org/10.14411/eje.2002.071
- Attié M, Baret S, Strasberg D (2005) Les insectes phytophages associés à des plantes exotiques envahissantes à l'île de La Réunion (Mascareignes). Revue d'Ecologie: La Terre et la Vie 60: 107–125
- Attié M, Bourgoin T, Veslot J, Soulier-Perkins A (2008) Patterns of trophic relationships between planthoppers (Hemiptera: Fulgoromorpha) and their host plants on the Mascarene Islands. Journal of Natural History 42 (23–24): 1591–1638
- Anufriev GA, Emeljanov AF (1988) Suborder Cicadinea (Auchenorrhyncha). pp. 12–496 in: PA Lehr (ed.) Keys to the identification of Insects of the Far East of the USSR. II. Homoptera and Heteroptera, Nauka Publishing House, Leningrad
- Bourgoin T (1987) A new interpretation of the homologies of the Hemiptera male genitalia, illustrated by the Tettigometridae (Hemiptera, Fulgoromorpha). Proceedings of the 6th Auchenorrhyncha meeting Turin, Italy 7–11 September 113–120
- Bourgoin T. (2023) FLOW (Fulgoromorpha Lists on The Web): a world knowledge base dedicated to Fulgoromorpha. Version 8, updated [Last update: 22 Feb. 2023]. Online at http://www.hemiptera-databases.org/flow/ [last accessed 27 Feb. 2023]
- Bourgoin T, Deiss V (1994) Sensory plate organs of the antenna in the Meenoplidae-Kinnaridae group (Hemiptera: Fulgoromorpha). International Journal of Insect Morphology and Embryology 23 (2): 159–168
- Bourgoin T, Wang RR, Asche M, Hoch H, Soulier-Perkins A, Stroinski A, Yap S, Szwedo J (2015) From micropterism to hyperpterism: recognition strategy and standardized homology-driven terminology of the fore wing venation patterns in planthoppers (Hemiptera: Fulgoromorpha). Zoomorphology 134 (1): 63–77. https://doi.org/10.1007/s00435-014-0243-6
- Ceotto P, Bourgoin T (2008) Insights into the phylogenetic relationships within Cixiidae (Hemiptera: Fulgoromorpha): cladistics analysis of a morphological dataset. Systematic Entomology 33: 484–500. https://doi.org/10.1111/j.1365-3113.2008.00426.x
- Distant WL (1906) The fauna of British India, including Ceylon and Burma. 3: 503 pp. Lt. Col. C. T. Birgham
- Emeljanov AF (2002) Contribution to classification and phylogeny of the family Cixiidae (Hemiptera, Fulgoromorpha). Denisia 4 (176): 103–112
- Emeljanov AF (2007) New and little-known taxa of the family Cixiidae (Homoptera, Fulgoroidea). Entomological Review 87: 287–308. https://doi.org/10.1134/S0013873807030062

- Erbe P, Hoch H (2004) Two new species of the Australian planthopper genus *Solonaima* Kirkaldy (Hemiptera: Fulgoromorpha: Cixiidae). Zootaxa 536: 1–7. https://doi.org/10.11646/zootaxa.536.1.1
- Fennah RG (1967) New and little known Fulgoroidea from South Africa (Homoptera). Annals of the Natal Museum 18 (3): 655–714
- Fennah RG (1969) Fulgoroidea (Homoptera) from New Caledonia and the Loyalty Islands. Pacific Insects Monography 21: 1–116
- Haglund CJE (1899) Beiträge zur Kenntnis der Insektenfauna von Kamerun. 4. Verzeichniss der von Yngve Sjöstedt im nordwestlichen Kamerungebeite eingesammelten Hemipteren. Ofversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar. Stockholm 56: 49–71
- Hoch H, Bonfils J, Reynaud B, Attié M (2003) First record of troglobitic Hemiptera (Fulgoromorpha: Cixiidae) from La Réunion Island. Annales de la Société Entomologique de France 39: 265–270
- Hoch H, Howarth FG (1989) Reductive evolutionary trends in two new cavernicolous species of a new Australian cixiid genus (Homoptera: Fulgoroidea). Systematic Entomology 14: 179– 196. https://doi.org/10.1111/j.1365-3113.1989.tb00276.x
- Holzinger WE, Emeljanov AF, Kammerlander I (2002) The family Cixiidae Spinola 1839 (Hemiptera: Fulgoromorpha) – a review. Denisia 4 (176): 113–138
- Holzinger WE, Kammerlander I, Nickel H (2003) The Auchenorrhyncha of Central Europe. Volume I: Fulgoromorpha, Cicadomorpha excl. Cicadellidae. Brill Leiden/Boston
- Holzinger WE, Löcker H, Löcker B (2008) Fulgoromorpha of Seychelles: a preliminary checklist. Bulletin of Insectology 61 (1): 121–122
- Jacobi A (1917) Die Zikadenfauna Madagascars und der Comoren. Pp. 519–552 in: Jacobi A. (ed.) Voeltzkow's Reise in Ostafrika in den Jahren 1903–1905 – Band III. E. Schweizbart'sche Verlagsbuchhandlung, Nägele & Dr. Sprösser. Stuttgart, Germany
- Jacobi A (1928) Rhynchota Homoptera. 1. Fulgoridae und Cercopidae in Results of Dr. E. Mjöberg's Swedish Scientific Expeditions to Australia 1910–1913. Arkiv for Zoologi. Utgifvet af K. Svenska Vetenskaps-akademien. Stockholm 19 (28): 1–50
- Kirkaldy GW (1906) Leafhoppers and their natural enemies. (Pt. IX Leafhoppers. Hemiptera). Bulletin. Hawaiian Sugar Planters' Association Experiment Station. Division of Entomology 1 (9): 271–479
- Knight WJ (1965) Techniques for use in the identification of leafhoppers (Homoptera, Cicadellidae). Entomologist's Gazette 16: 129–136
- Lallemand V, Synave H (1954) Homoptères nouveaux de Madagascar. Le Naturaliste Malgache 6: 79–82
- Löcker B, Fletcher MJ, Gurr GM (2007) Revision of the genus *Innobindus* Jacobi (Hemiptera: Fulgoromorpha: Cixiidae) with the description of six new species and comments on other Australian Brixiini genera. Australian Journal of Entomology 46 (1): 45–55
- Löcker B, Fletcher MJ, Gurr GM (2010) Taxonomic revision of the Australian Eucarpiini (Hemiptera: Fulgoromorpha: Cixiidae) with the description of nine new species. Zootaxa 2425: 1–31. https://doi.org/10.11646/zootaxa.2425.1.1
- Metcalf ZP (1938) The Fulgorina of Barro Colorado and other parts of Panama. Bulletin of the Museum of Comparative Zoology at Harvard College. Cambridge, Mass 82: 277–423
- Muir FAG (1922) New Malayan Cixiidae (Homoptera). Philippine Journal of Science. Manila 20 (1): 111–121

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- Paulian R (1953) Une campagne spéléologique dans la réserve naturelle de Namoroka. Naturaliste malgache 5 (1): 19–28
- Romani R, Stacconi MVR, Riolo P, Isidoro N (2009) The sensory structures of the antennal flagellum in *Hyalesthes obsoletus* (Hemiptera: Fulgoromorpha: Ciixidae): A functional reduction? Arthropod Structure & Development 38: 473–483. https://doi.org/10.1016/j.asd.2009.08.002
- Signoret V (1860) Faune des hémiptères de Madagascar. 1ère partie. Homoptères. Annales de la Société Entomologique de France 8: 177–206
- Soulier-Perkins A, Ouvrard D, Hoch H, Bourgoin T (2015) Singing in the Namoroka Caves, First Record In Situ for a Cave Dwelling Insect: *Typhlobrixia namorokensis* (Hemiptera, Fulgoromorpha, Cixiidae). Journal of Insect Behaviour 28: 704–721. https://doi.org/10.1007/s10905-015-9531-3
- Spinola M (1839) Essai sur les Fulgorelles, sous-tribu de la tribu des Cicadaires, ordre des Rhyngotes. Annales de la Société Entomologique de France. Paris 8: 133–337
- Stål C (1856) Om Derbides med tre oceller. Ofversigt af Kongliga Svenska Vetenskaps-Akademiens Förhandlingar. Stockholm 13: 161–164
- Stål C (1866) Hemiptera Homoptera Latr. Hemiptera Africana 4: 1–276
- Synave H (1952) Description de deux Cixiidae nouveaux appartenant au genre *Achaemenes* Stål, et note préliminaire sur un Cixiide troglobie. Bulletin de l'Institut Royal des Sciences Naturelles de Belgique 28 (60): 1–8
- Synave H (1953) Un Cixiide troglobie découvert dans les galeries souterraines du système de Namoroka (Hemiptera – Homoptera). Naturaliste malgache 5 (2): 175–179
- Synave H (1956) Les Cixiidae de Madagascar (Hemiptera Homoptera). Mémoires de l'Institut des Sciences de Madagascar (Ser. E) 7: 167–196
- Synave H (1965) Contribution à la connaissance des Cixiidae de Madagascar, Maurice et la Reunion (Homoptera-Fulgoroidea). Bulletin de l'Institut Royal des Sciences Naturelles de Belgique 41 (27): 1–57

- Synave H (1969) Note sur le genre *Taosa* Distant (Dictyopharidae) et sur *Harmosa bivulneratum* Fennah (Eurybrachidae) (Homoptera Fulgoroidea). Bulletin de l'Institut Royal des Sciences Naturelles de Belgique 45 (8): 1–17
- Synave H (1979) Description d'espèces nouvelles appartenant aux familles: Cercopidae, Cixiidae, Derbidae, Dictyopharidae et Tropiduchidae (Homoptera). Bulletin du Musée Royal d'Histoire Naturelle de Belgique. Bruxelles 51 (6): 1–31
- Szwedo J (2004) *Autrimpus sambiorum* gen. *et* sp. nov. from Eocene baltic amber and notes on Mnemosynini stat. nov. (Hemiptera: Fulgoroidea: Cixiidae). Annales Zoologici 54: 567–578
- Tsaur S-Ch, Hsu T-Ch (2003) The Cixiidae of Taiwan, Part VII: Tribe Pintaliini (Hemiptera: Fulgoroidea). Zoological Studies 42 (3): 431–443
- Van Stalle J (1984) Les Cixiides de la Forêt de Taï (Côte-d'Ivoire): description de neuf espèces nouvelles (Homoptera, Fulgoroidea). Revue Française d'Entomologie. Paris 6 (3): 137–146
- Van Stalle J (1987) A review of the genus Caffrocixius Fennah, 1967 with description of four new species (Homoptera, Cixiidae). Mitteilungen der Schweizerischen Entomologischen Gesellschaft 60: 347–353
- Walker F (1857) Catalogue of the Homopterous insects collected at Sarawak, Borneo, by Mr. AR Wallace, with descriptions of new species. Journal of the Proceedings of the Linnean Society. London 1: 141–17
- Walker F (1870) Catalogue of the Homopterous insects collected in the Indian Archipelago by Mr. AR Wallace, with descriptions of new species. Zoological Journal of the Linnean Society. London 10: 82–193
- Wang RR, Liu JJ, Li XY, Liang AP, Bourgoin T (2018) Relating antennal sensilla diversity and possible species behaviour in the planthopper pest *Lycorma delicatula* (Hemiptera: Fulgoromorpha: Fulgoridae). Plos ONE 13: e0194995. https://doi.org/10.1371/journal.pone.0194995
- Williams JR (1975) Cixiidae (Fulgoroidea: Homoptera) from Réunion Island. Journal of Natural History 9: 669–680